

Novel Adsorbents from Acrocomia Totai Leaves for Caffeine Removal: A Sustainable Approach

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Abstract

This study focused on the production of activated hydrochars from Acrocomia Totai leaves. The hydrochars were synthesized through hydrocarbonization with KOH in a ratio of 1:5 (m/m). The specific surface area (SBET) was determined to be 2.132 m².g⁻¹ for HDML-ATV, 1726 m².g⁻¹ for HME4h-ATV and 1.735 m².g⁻¹ for HME48h-ATV. Regarding the maximum amount adsorbed, the hydrochars gave the following results: 411.78 mg g⁻¹ for HMDL-ATV, 344.52 mg g⁻¹ for HME4h-ATV, and 303.43mgg-1 for HME48h-ATV. The results of this study demonstrate the potential of using Acrocomia Totai leaves for the production of activated hydrochars with large surface areas, which can be used for various applications, including the adsorption of caffeine (CFN).

Keywords: Hydrochars; Caffeine adsorption; Green chemistry; Acrocomia totai leaves.

1. Introduction

The macauba palm, native to Brazil, is known for its versatile uses[1]. Its leaves are used for biocomposite materials, while the seeds are used for biofuel production. The production of porous materials from macauba is of particular interest, as it offers a sustainable alternative for various applications[2]. The leaves and seeds are rich in cellulose and lignin, making them suitable for the production of porous materials such as activated carbon and biochar. These materials have a wide range of uses, including water filtration, air purification, and as a substrate for catalytic reactions. The use of macauba for the production of porous materials aligns with the growing demand for sustainable and eco-friendly alternatives in various industries.

2. Synthesis of hydrochars

This study focused on Macaúba palm trees from Moreira Sales, Paraná, Brazil (latitude -24° 3' 23.64", longitude -53° 3' 26.32"), selected as the raw material for hydrochar production. Three samples were collected: one from fallen burlap and two from lipid-extracted Macaúba using a Soxhlet apparatus for either 4 or 48 [3].

Hydrochar produced was via hydrocarbonization by placing 5.00 g. of each sample in a Teflon®-lined stainless steel autoclave with 37 mL of deionized water, heated to 190°C for 48 hours. The resulting hydrochars were dried at 70°C for 12 hours and labeled as HMDL (burlap-derived), (4-hour lipid HME4h extraction), HME48h (48-hour lipid and extraction).

2.1 Chemical activation of hydrochars.

Samples were chemically activated with KOH (1:5 ratio), mixed with deionized water, stirred, and heated to 800°C in argon gas. Post-activation, samples were washed with deionized water and 0.1 M HCl, then dried at 100°C for 12 hours.

2.2 Characterization of hydrochars and activated hydrochars

Nitrogen physisorption isotherms at 77 K for activated hydrochars were measured using a QuantaChrome Nova 1200e, with BET surface



areas (S_{BET}) from 0.05–0.20 relative pressure. Total pore volume (V_T) was determined at p/p_o = 0.99. FTIR-ATR and Raman spectra were recorded using Bruker spectrophotometers, while SEM evaluated morphological features.

2.4 Adsorption studies

Adsorption studies used 20.0 mL CFN solutions and 10 mg activated hydrochars in 50 mL flasks, shaken at 70 rpm and 25°C. CFN concentrations were measured with a Cary 50 spectrophotometer after 0.44 μ m vacuum filtration. The effects of pH (3.0-11.0), time (0-480 min), and adsorption isotherms (100-500 mg/L CFN, pH 7.0) were evaluated using Langmuir, Freundlich, Dubinin-Radushkevich, and Redlich-Peterson models.

The maximum CFN amounts adsorbed onto activated hydrochars, qe (efect of pH (3.0 - 11.0), adsorption isotherms and temperature effect (25. 45 e 55°C) and the CFN amounts adsorbed at time t, q_t, were determined from Equation 1.

$$\mathbf{q}_{t} = \mathbf{q}_{e} = \frac{\left(\mathbf{C}_{0} - \mathbf{C}_{f}\right)\mathbf{v}}{\mathbf{w}}$$
(1)

Isothermal and kinetic models were fitted to experimental data using a nonlinear equations. The data were processed using Origin[®] 8.5 software. the normalized standard deviations (Δ_{qe}) (Equation 2) and the correlation coefficiente (R²) were used to compare the applicability of the CFN adsorption data to the proposed model.

$$\Delta q_e(\%) = 100 \sqrt{\sum \frac{\left[\frac{q_{e, \exp} - q_{e, cal}}{q_{e, \exp}}\right]^2}{N - 1}}$$
(2)

Where $q_{e,exp}$ and $q_{e,cal}$ are experimental and calculated values of the adsorbed amounts, respectively, and N represents the number of experiments.

3. Results and discussions

3.1 Characterization of HM-ATV

The textural properties of HMDL-ATV, HME4h-ATV, and HME48h-ATV are shown in

Table 1. HMDL-ATV has a high S_{BET} (2132 m²g⁻¹) due to the burlap's high carbon content. HME4h-ATV and HME48h-ATV, with extracted compounds where carbon is the major element, have lower S_{BET} values than HMDL-ATV. Additionally, the samples exhibit a high mesopore/micropore volume ratio and an average pore diameter of 2.25 nm. HME4h-ATV and HME48h-ATV have high S_{BET} values of 1726 m²g⁻¹ and 1735 m²g⁻¹, respectively.

Table 1. Textural properties of HMDL-A TV, HME4h-ATV and HME4h-ATV.

Sample	S _{BET}	V _T	V _{micro}	V _{meso}	D _P
HDML-ATV	2132	1120	0.801	0.319	2.25
HME4h- ATV	1726	0.996	0.672	0.324	3.40
HME48h- ATV	1735	0.974	0.692	0.282	2.24

 $\overline{S_{BET}(m^2g^{-1})}; V_T(cm^3g^{-1}); V_{micro}(cm^3g^{-1}); Vmeso(cm^3g^{-1}); Dp(nm).$

The SEM images of HMDL-ATV, HME4h-ATV, and HME48h-ATV are shown in Figure 2A, B, and C reveal a dense, irregular morphology without cavities. In contrast, the SEM images of the activated hydrochars display numerous cavities of various structures, formed due to the KOH chemical activation of Macaúba leaves. These cavities on the surface of the activated hydrochars may be responsible for CFN adsorption.



Figure 2- SEM images A- HMDL-ATV, B- HME4h-ATV and HME48h-ATV

Figure 3A shows the FTIR spectra of activated hydrochars, indicating different chemical bonds and functional groups. All materials exhibit similar profiles: -OH stretching in hydroxyl and carboxyl groups (~3600 cm⁻¹), C-H stretching in aliphatic and aromatic structures (~3000 cm⁻¹), and C=C presence (~1650 cm⁻¹). Hydrochars show bands at 1530, 1142, 934, and 521 cm⁻¹, while activated hydrochars reveal new bands at 1340 and 707 cm⁻¹. These findings align with previous studies [4,5].



The Raman spectra of activated hydrochar and hydrochar (Figure 3B) show two bands: D at 1348 cm⁻¹ and G at 1591 cm⁻¹. The D band indicates defects, while the G band represents ordered sp² carbon. The graphitization degree, determined by the ID/IG ratio, is higher in hydrochar (ID/IG < 0.89) than in activated hydrochar (ID/IG > 0.97), indicating reduced graphitization post-activation.



Figure 3 - A- FTIR spectra and B- Raman scattering of activated hydrochars

3.2 Adsorption studies

3.2.1 Adsorption kinetics

Figure 4 shows CFN adsorption kinetics on three activated hydrochars, with rapid increase in the first 120 min, reaching equilibrium at 480 min. High q_{tq} values are seen early. Non-linear kinetic models (pseudo-first order, pseudo-second order, and Elovich) fit the experimental data. Table 3 presents the kinetic parameters.



Figure 4 - Adsorption kinetics of CFN on A - HMDL-ATV, B – HME4h-ATV and C – HME48h-ATV

Table 2 shows that the pseudo-second-order model best correlated with the experimental data for HMDL-ATV and HME4h-ATV, while the Elovich model fit HME48h-ATV data best. These models had high R² (0.9825, 0.9844, 0.9813) and low standard deviations (Δ_{qe} = 4.43, 6.79, 8.94). Initial adsorption rates confirm quick CFN adsorption, with HMDL-ATV > HME4h-ATV > HME48h-ATV, reflecting their S_{BET} and micropore volume. These properties effectively retain caffeine molecules (dimensions in nm: 1.06 length, 0.85 width, 0.45 thickness).

Kinetic parameter					
Sample	Pseudo-First Order				
(I)	qe=464.50	k1=0.41	h ₀ =181.15	R ² =0.9590	
(11)	qe=391.35	k1=0.28	h ₀ =109.57	R ² =0.9535	
(111)	q _e =349.55	k1=0.31	h ₀ =108.36	R ² =0.9312	
	Pseu	do Second	Order		
(I)	qe=457.09	k ₂ =0.001	h ₀ =334.29	R ² =0.9825	
		6			
(11)	qe=411.18	$k_2 = 0.001$	h ₀ =185.97	$R^2 = 0.9844$	
		1			
(111)	qe=366.94	$k_2 = 0.001$	$h_0 = 175.13$	R ² =0.9744	
		3			
Elovich					
(I)	$\alpha = 2.64 10^6$	β=0.036 R ² =0.9		R ² =0.9806	
(11)	$\alpha = 3.04 10^5$	β=0.028		R ² =0.9645	
(111)	α= 3.38 10 ⁵	β=0.032		R ² =0.9813	
15 4	1 1 4 1	. 1 .	1 . 1	1 1 2 0	

Table 2. Kinetic paran	neters of CFN adsorption	on
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 $\begin{array}{l} q_{e} \ (mg \ g^{-1}); \ k_{1} = \ min^{-1}; \ h_{0} = \ mg \ g^{-1} \ min^{-1}; \ k_{2} = \ g \ mg^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-2}; \beta = \ mg \ g^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ min^{-1}; \ \alpha = \ g \ mg^{-1} \ mg^{-1}; \ mg^{-1};$

3.2.2 Adsorption isotherms

The isotherms of caffeine adsorption onto the synthesized activated hydrochars were investigated and are shown in Figure 5. Non-linear isotherm models (Langmuir, Freundlich, Dubinin-Radushkevich, and Redlich-Peterson) and their parameters are displayed in Table 4.



Figure 5 - Adsorption isotherms of CFN on A - HMDL-ATV, B – HME4h-ATV and C – HME48h-ATV

From the adsorption isotherm , the maximum CFN adsorption capacities onto activated hydrochars were determined: 411.78 mg g⁻¹ for HMDL-ATV, 344.52 mg g⁻¹ for HME4h-ATV, and 303.43mgg⁻¹ for HME48h-ATV [6]. Langmuir and Redlich-Peterson models showed the best correlation, with high R² values and low Δ_{qe} The Langmuir model estimated close to experimental monolayer adsorption capacities (q_{max}), suggesting homogeneous adsorption sites.



Sample		Isot	therms parame	ters		
Langmuir						
(I)	q _{max} =464.50	$k_{L}=0.1$	$6 \qquad \Delta q_e = 3$.77	R ² =0.9907	
(II)	q _{max} =356.98	k _L =0.1	1 $\Delta q_e = 5$.92	R ² =0.9907	
(III)	q _{max} =300.13	k _L =0.16	$\Delta q_e = 4$.63	R ² =0.9849	
		Fre	undlich			
(I)	k _F =165.00	n= 5.9	9 $\Delta q_e = 10.$	84	$R^2 = 0.9627$	
(II)	k _F =173.35	n= 7.8	8 $\Delta q_e = 12.$	74	$R^2 = 0.8845$	
(III)	$k_F = 134.74$	n= 6.8	$6 \qquad \Delta q_e = 4.3$	9	R ² =0.9905	
Dubinin-Raduchkevich						
(I)	$q_m = 382.24$	$k_{DR} =$	E=373.71	$\Delta q_e =$	$R^2 =$	
		3.58E ⁻⁰⁶		10.34	0.6541	
(II)	$q_m = 328.08$	$k_{DR} =$	E=298.15	$\Delta q_e =$	$R^2 =$	
		5.98E ⁻⁰⁶		12.73	0.9541	
(III)	$q_m = 284.53$	k _{DR} =	E=320.42	$\Delta q_e =$	$R^2 =$	
		5.98E ⁻⁰⁶		8.15	0.9534	
Redlich-Peterson						
(I)	$a_{RP} = 77.97$	$b_{RP} =$	g=0.9701	$\Delta q_e =$	$R^2 =$	
		0.224		24.94	0.9902	
(II)	a _{RP} =587.81	$b_{RP} =$	g=0.931	$\Delta q_e =$	$R^2 =$	
		0.235		23.61	0.9918	
(III)	a _{RP} =105.41	$b_{RP} =$	g=0.910	$\Delta q_e =$	$R^2 =$	
		0.572		16.05	0.9945	
$q_{max} = (mg g^{-1}); k_L = (L mg^{-1}); k_F = [mg g^{-1} (mg L^{-1)-1/n}], qm = (mg g^{-1}); k_{DR} = (mol^2 kJ^{-2});$						

Table 2. Isotherms parameters of CFN adsorption

 $\begin{array}{l} q_{max} = (mg \ g^{-1}); \ k_L = (L \ mg^{-1}); \ k_{F^{-1}} \ [mg \ g^{-1} (mg \ L^{-1p+lm}], \ qm = (mg \ g^{-1}); \ k_{DR} = (mol^2 \ kJ^{-2}); \\ E \ (KJ \ mol^{-1}); \ a_{RP} = (L \ mg^{-1})^{-g}; \ b_{RP} = L \ g^{-1}; \ \textbf{(I) HDML-ATV; (II) HME4h-ATV; (III)} \\ \textbf{HME48h-ATV} \end{array}$

The Redlich-Peterson model, a three-parameter empirical model combining Langmuir and Freundlich isotherms, shows parameter g close to 1, indicating behavior similar to the Langmuir model for CFN adsorption (Khasri et al., 2018; Wan et al., 2018). The Dubinin-Radushkevich model exhibits high R^2 values (>0.92), suitable for understanding adsorption free energies (E). All three activated hydrochars show E values >8.00 kJ mol⁻¹, suggesting predominant chemisorption.

4 Conclusion

In conclusion, HMDL-ATV, HME4h-ATV, and HME48h-ATV demonstrated varying caffeine adsorption capacities, reflecting their distinct surface properties and suitability for environmental remediation applications.

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